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(21) International Application Number: PCT/NL96/00198 (22) International Filing Date: 8 May 1996 (08.05.96) (30) Priority Data: 95201197.1 9 May 1995 (09.05.95) EP (34) Countries for which the regional or international application was filed: NL et al. (71) Applicant (for all designated States except US): KREATECH BIOTECHNOLOGY B.V. [NL/NL]; Kelenbergweg 3, NL-1101 EZ Amsterdam (NL). (72) Inventors; and (75) Inventors/Applicants (for US only): HOUTHOFF, Hendrik, Jan [NL/NL]; Schellingwouderdijk 125, NL-1023 NB Amsterdam (NL). REEDIJK, Jan [NL/NL]; Anthoni Duycklaan 4, NL-2334 CD Leiden (NL). JELSMA, Tinka [NL/NL]; Kerseboomstraat 30, NL-1326 DG Almere (NL). VAN ES, Remco, Maria [NL/NL]; Henneland 25, NL-1541 NR Koog a/d Zaan (NL). VAN DEN BERG, Franciscus, Michiel [NL/NL]; Klaterbos 75, NL-2134 JB Hoofddorp (NL). LEMPERS, Edwin, Leo, Mario [NL/NL]; Vogelzand 2231, NL-1788 GB Julianadorp (NL). BLOEMINK, Marieke, Johanna [NL/NL]; Hofbroeckerlaan 31, NL-2341 LM Oegstgeest (NL).		(74) Agent: SMULDERS, Th., A., H., J.; Vereenigde Octrooibureaux, Nieuwe Parklaan 97, NL-2587 BN The Hague (NL). (81) Designated States: AL, AM, AT, AU, AZ, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, US, UZ, VN, ARIPO patent (KE, LS, MW, SD, SZ, UG), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i>
(54) Title: METHODS FOR THE PRODUCTION OF PLATINUM-BASED LINKERS BETWEEN LABELS AND BIO-ORGANIC MOLECULES, FOR LABELLING BIO-ORGANIC MOLECULES, FOR DETECTING BIOLOGICAL SUBSTANCES OF INTEREST AND DIAGNOSTIC TEST KITS (57) Abstract <p>The present invention provides improved methods of producing platinum compounds, which are very suitable for producing labelled substances, which can be used to detect specific molecules of interest. The platinum coordination compounds have two reactive groups of which one is replaced by a label and the other one can be replaced by a substance to be labelled. Production of labelled substances is very much improved by selection of the right starting materials and producing the right intermediates. The efficiency of labelling is very much improved, thereby enabling the production of labelling kits which are also a part of the present invention.</p>		

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Title: Methods for the production of platinum-based linkers between labels and bio-organic molecules, for labelling bio-organic molecules, for detecting biological substances of interest and diagnostic test kits.

The invention relates to methods for the production of linkers (linking moieties between labels and bio-organic molecules, which linkers are based on platinum compounds. Platinum (coordination) compounds have been considered
5 interesting molecules for a very long time. For a review of these compounds and their uses we refer to Reedijk et al. (Structure and Bonding 67, p.53-89, 1987). Especially Cis-platinum has received a lot of attention as a possible anti-tumour drug. This anti-tumour reactivity of platinum compounds
10 originates from their having at least two reactive groups (preferably cis-oriented towards each other), which make it possible to cross-link DNA molecules, thereby inhibiting the replication of these DNA molecules.

A different use of platinum (coordination) compounds has
15 been disclosed in PCT application (WO92/01699) wherein a platinum compound having two reactive moieties (denominated as leaving groups therein) is reacted with a fluorescein to obtain a labelled platinum compound having only one reactive moiety which can bind (non-covalently) to a nucleic acid,
20 preferably at the N-7 position of a guanine residue. The present invention provides improved methods of preparing such platinum compounds which are suitable for use as linkers in such labelling substances, improved methods of preparing labelled substances and the use of such substances in
25 preparing diagnostic test kits.

In WO92/01699 the starting compounds disclosed are platinum(II)(ethylenediamine)dichloride or [platinum(II)(ethylenediamine)(Me₂SO)Cl]⁺Cl⁻.

Both compounds have certain disadvantages for use as a linker. The first one can be obtained commercially, the second one (the preferred one) must be synthesized as the chloride salt. In the dichloride compound, the Cl^- ions are less readily substituted by a marker group or a guanine group, respectively. Moreover, since the two Cl^- ions have about the same reactivity, it happens that both ions are substituted by a label, leaving no reactive moiety for coupling the labelled substance to a bio-molecule of interest. It is for that reason that compounds having different reactive moieties are very much preferred for use in labelling. This is true for the second compound, however, the total DNA labelling for this compounds will be appreciably longer, up to several hours instead of several minutes, making the product less suitable for routine use.

We have now found a very simple and reliable method to produce symmetrical linkers including the first one just mentioned and a number of novel ones, through selection of suitable starting compounds of the formula PtE_4 wherein E is an electronegative group, preferably a halogen or NO_3^- or SO_3^- . The reaction, which is described in the examples, of these starting compounds with ethylenediamine is very simple and efficient. Moreover, this reaction leads to very suitable symmetric intermediate compounds for producing labelled substances. The advantages of working with these starting compounds for producing labelled substances have never been disclosed and these starting compounds have never been used for this purpose. A major advantage of using these compounds is that when a stabilizing bridge for the resulting platinum compound has to be attached that no blocking reagents have to be employed. Another advantage is that the resulting intermediate compounds can again be labelled without the use of blocking agents. Therefore steps removing blocking agents can be eliminated completely. Furthermore the yields of these reactions are very high. Another advantage of the use of these symmetrical starting compounds is that no mixtures of different resulting compounds can be formed, which may

interfere with the following reaction and reduce yield or require extra separation steps. A very suitable intermediate compound according to the invention is platinum(II)(ethylenediamine)(NO₃)₂. Surprisingly, hardly any double labelling occurs with these compounds, which is in contrast with the dichloride compound of WO92/01699. This substance can very easily be provided with a suitable labelling group, resulting in a labelling substance which can still, through substitution of the remaining NO₃ group be linked to a bio-organic molecule to be labelled or detected, which reaction proceeds much faster than with the known platinum compounds. Furthermore the methods for producing this compound and the resulting compound do not involve highly toxic substances such as DMSO.

The intermediate compounds can be labelled with any suitable label (also known as marker). These labels may be radioactive labels, enzymes (which need reaction with a substrate to be detected), specific binding pairs components such as avidin, streptavidin or biotin, biocytin, iminobiotin, colloidal dye substances, fluorochromes (rhodamin, etc.), reducing substances (eosin, erythrosin, etc.), (coloured) latex sols, digoxigenin, metal sols or other particulate sols (selenium, carbon and the like), dansyl lysin, antibodies, protein A, protein G, etc. These labels may be attached to the linkers directly or through spacer arms (preferably polylysine). Because the labelling of nucleic acids and proteins is so easy, it is possible to sell the linker-label compounds separately in a kit so that any user can produce the labelled substances easily, or can attach by the same procedure many different labels of choice to the substances. This allows for multiple detections in one assay. Furthermore the known advantages of the platinum labelling compounds (from WO92/01699 for instance) are of course also obtained with the present methods and compounds. Another advantage of the platinum compounds is that they can be detected more or less directly by using the platinum as a nucleus for depositing silver or other metal crystals.

Almost every bio-organic molecule which contains an accessible S (sulphur) or an N (nitrogen) can be labelled with the platinum compounds. A very suitable bio-organic molecule to be labelled with the labelling substance according to the invention is nucleic acid (nucleotides, oligonucleotides, DNA, RNA, homo duplexes, heteroduplexes, or multiplexes). The platinum binds very easily (non-covalently) to the N-7 position of guanine residues. This way DNA or RNA molecules, be it single stranded or otherwise can be easily detected, but it also allows for the production of probes for hybridization techniques wherein unlabelled DNA/RNA molecules hybridize to the labelled probe. The platinum compounds do hardly interfere with the hybridization, if at all. Also, this technique obviates the use of modified nucleotides in preparing probes. However, proteins, peptides and other bio-organic molecules can also be labelled with the labelling substances according to the invention.

The platinum compounds according to the invention are also very suitable for attaching bio-organic molecules to solid surfaces such as nitrocellulose, nylon filters or microtiter plates.

Nucleotides modified in accordance with the practices of this invention and oligo- and polynucleotides into which the modified nucleotides have been incorporated or oligo- and polynucleotides that have been directly modified using these novel platinum compounds may be used as probes in biomedical research, clinical diagnostics and recombinant DNA technology.

The wide variety of utilities are based upon the ability of the platinum compounds to form stable complexes with polypeptides which in turn can be detected either by means of detectable moieties which are attached to or which interact with the polypeptide. Some uses include detecting and identifying nucleic acid-containing etiological agents, e.g. bacteria and viruses; screening bacteria for antibiotic resistance; screening animals for genetic disorders in relation to pharmaceutical effects; diagnosing genetic

disorders, e.g. trisomy 21, sickle cell anemia: chromosomal karyotyping: and identifying tumour cells.

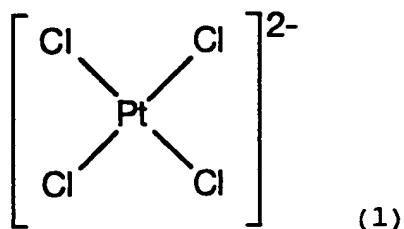
Other advantages and embodiments of the invention will become clear from the following experimental part.

EXPERIMENTAL

SYNTHESIS OF INTERMEDIATE PLATINUM COMPOUNDS

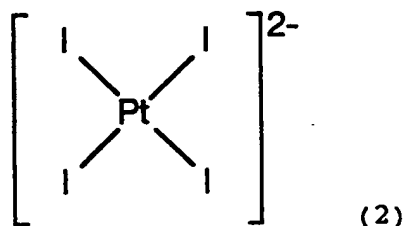
5 These compounds may be prepared by a process which involves:

(a) reacting a compound in which the anion has the structure:



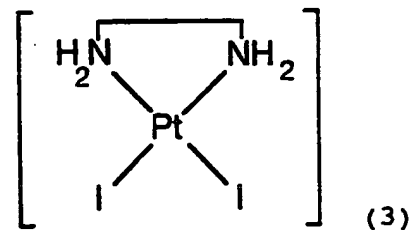
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with KI in a suitable solvent under suitable conditions so as to form a iodated platinum compound wherein the anion has the structure:

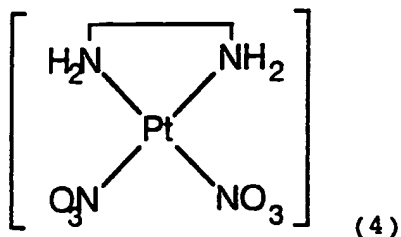


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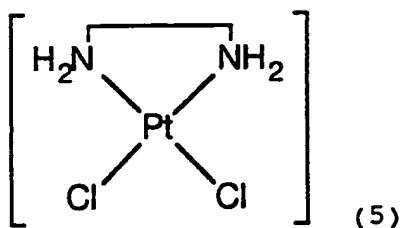
(b) reacting said iodated platinum compound (2) with ethylenediamine in a suitable solvent so as to form a diethylenediamine iodated platinum compound as represented by
20 the formula $\text{Pt}(\text{en})\text{I}_2$ and which has the structure:



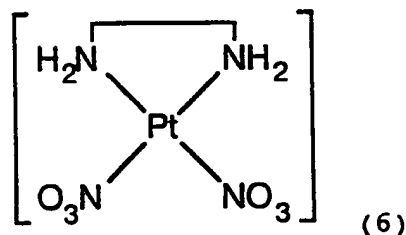
(c) reacting said compound (3) with AgNO_3 , the reaction being carried out in a suitable solvent, under suitable conditions so as to form a compound having the structure:



(d) reacting said compound (4) with KCl in a suitable solvent under suitable conditions so as to form a compound having the structure:



(e) reacting said compound (5) with AgNO_3 in a suitable solvent, under suitable conditions so as to form a compound having the structure:



(f) recovering said compound (6) as modified platinum starting compound for the synthesis of hapten-bound $\text{Pt}(\text{en})$ compounds for use as DNA and/or RNA label.

Compounds (4) and (6) are of course the same, both reactions preparing them have been given as alternatives.

Example 1

5

Preparation of Pt(en)-diamine starting material.

Preparation of Platinumethylenediamine(NO₃)₂: starting material.

10 Pt(en)(NO₃)₂

All reactions are performed in the dark.

Dissolve 1 gram Potassium tetrachloroplatinate(II), K₂PtCl₄ (2.4 mmol, Sigma) in 50 ml millipore (filtered water) and stir at room temperature. Add 10 equivalents of Potassium iodide, KI (24 mmol, 3.999 g, Sigma). The colour of the solution will immediately turn from orange into dark red (K₂PtI₄), stir for 5 minutes.

Add one equivalent ethylenediamine (2.4 mmol, 160.8743 µl, Merck 11=0.9 kg) after diluting 161 µl ethylenediamine in 5 ml millipore very slowly to the platinum solution, mix this solution for 1 hour at room temperature.

A yellow/brown precipitate, Pt(en)I₂, will be formed and the liquid standing above should be clear.

25 Filter the solution through a 1.0 µm membrane filter (Schleicher&Schuell), wash the precipitate with millipore, ethanol and diethylether (in this order). Dry the Pt(en)I₂ for at least 4 hours in a vacuum dry-oven at 37°C.

30 Weigh the dried Pt(en)I₂ (~1.07 g) and suspend it in 45 ml millipore/5 ml aceton, the solution will be cloudy. Add 1.95 equivalent of AgNO₃ (M = 169.9, Sigma). Stir the reaction overnight at room temperature.

35 Filter the solution through a 1.0 µm membrane filter, the precipitate is Silver iodide, AgI, the filtrate should be clear.

Add to 0.5 ml of the filtrate, containing Pt(en)(NO₃)₂, an excess of KCl or NaCl and make sure that no white precipitate is formed immediately after adding the excess of KCl or NaCl. If no white precipitate (only a yellow one) is formed then add

an excess of KCl or NaCl to the entire filtrate. After the yellow precipitate is formed filter the solution and wash the precipitate (Pt(en)Cl₂) with millipore, ethanol and diether.

- 5 Dry the precipitate for at least 4 hours in a vacuum
dryoven at 37°C.

Weigh the dry Pt(en)Cl₂ (M=326,1), and suspend it in 45 ml millipore/5 ml aceton and stir the cloudy suspension. Add 1.95 equivalent AgNO₃ and stir the solution overnight at room temperature. The colour of the solution will become white, due
10 to the formation of AgCl.

Filter the solution in the dark and evaporate the filtrate to remove the aceton by rotation evaporation untill 25 ml of the filtrate is left. The filtrate is then
freezedried. The product is checked by NMR or Infrared
15 Absorption Spectroscopy. An element analysis is given below.

Table I

20	Pt(en)(NO ₃) ₂	M _w = 379.1	20/1 batch		
	A	B	Δ A-B	Theor.	
25	Sample 2 :	5.79 % C	5.15 % C	-0.86 %	6.33 %
		2.03 % H	1.81 % H	+0.19 %	2.11 %
		12.1 % N	13.3 % N	-2.07 %	14.77 %
		48.2 % Pt	48.4 % Pt	-3.16 %	51.46 %
		23.5 % O	23.1 % O	-2.02 %	25.32 %
		<0.2 % Na			

There was not sufficient material for Cl-test.

V₂O₅ was added for the C,H-, N- and S-combustions.

Table II

Compound	Reqd.	Theory	Found (A)%	Ref.	Found (B)%	Ref.
ULS 95 RM005						
Probable	C	6.3	6.47	F2576		
Structure	H	2.1	2.11	"		
[Pt(en)(NO ₃) ₂	N	14.8	14.48	"		
	O					
	S					
	Hal					
	Metal					

Example 2

Preparation of immunologically detectable hapten-
Pt(en)compounds

Preparation of platinum ethylene diamine-ε-(6-
 (Biotinoyl)amino) hexanoyl-L-lysine-nitrate
 [Pt(en)(Biocytin-X)(NO₃)]NO₃.

All reactions are performed in the dark.

Dissolve 31.6 mg biocytin-X (0.065 mmol, Molecular
 Probes, Inc, USA) in 15 ml millipore in the dark and heat the
 solution slightly (up to a maximum 40°C) untill dissolving and
 adjust the pH to 7-8 with 0.4 M NaOH. Dissolve 23.4 mg
 Pt(en)(NO₃)₂ (M=379.1, 0.062 mmol)(see example 1) in 10 ml
 millipore in the dark and heat the solution slightly (up to a
 maximum of 50°C) until Pt(en)(NO₃)₂ is completely dissolved.

When both reagents are completely dissolved add the
 biocytin solution to the platinum solution and let them react

for 210 minutes at room temperature in the dark. The pH of the solution has to be 7.0, so the pH is monitored during the reaction and either H₂O or HNO₃ is added if necessary.

Isolate the product [Pt(en)(biocytin-X)(NO₃)]NO₃ by means
5 of freeze drying.

Stability of the product is ensured by the binding of Pt not only to the amino group of biocytin-X, but also to the carboxylate group of biocytin-X. Due to the latter binding kinetics of the platinum compound a chelate ring form is
10 created which will account for the stability of the product once it is dissolved in water, because no polymerisation reactions can occur.

An element analysis of [Pt(en)(biocytin-X)(NO₃)]NO₃ is given below.

15

Table III

BIO-ULS [Pt(en)BIO]NO ₃ M _w = 689,6				
	A	B	Δ A-B	Theor.
20 Sample 1 :	31.49 % C	31.42 % C	-0.064 %	31.50 %
	5.63 % H	5.40 % H	+0.374 %	5.89 %
	13.2 % N	13.0 % N	-1.24 %	14.34 %
	18.3 % Pt	18.5 % Pt	- 9.95 %	28.35 %
25	3.65 % S	3.67 % S	-1.0 %	4.66 %
	19.2 % O	19.0 % O	-2.826 %	21.926 %
	3.23 % Na			

There was not sufficient material for Cl-test.

Example 3**Preparation of Platinum ethylenediamine-Digoxigenin^R-L-lysine-nitrate.**

5 **[Pt(en)(Digoxigenin)(NO₃)](NO₃).**

Dissolve Pt(en)(NO₃)₂ (0.062 mmol, 23,65 mg) in 20 ml methanol and heat to 50°C until Pt(en)(NO₃)₂ is completely dissolved.

10 Dissolve digoxigenin-L-lysine (45,3 mg) in 20 ml methanol, heat until all solid material is dissolved.

Add the two solutions together and react for at least 3 hours. Isolate the end product by rotation evaporation. Digoxigenin^R is a registered trademark of Boehringer Mannheim.

15

Example 4**Preparation of fluorescent Platinum(en) compounds**

20 **Preparation of Platinum ethylenediamine-Fluoresceinamine-nitrate.**
[Pt(en)(Fluoresceinamine)(NO₃)]NO₃.

All reactions are performed in the dark.

25 Dissolve Pt(en)(NO₃)₂ (0.25 mmol, 94,8 mg) in 10 ml methanol/2 ml water, by heating until dissolving.

Dissolve fluoresceinamine (0.27 mmol, 95,5 mg, Sigma) in 10 ml methanol.

30 Add the fluoresceinamine solution to the platinum solution, an immediate change of colour is observed.

Run the reaction for at least 3 hours.

Isolate the crystalline end product by rotation evaporation, followed by freeze drying.

Example 5

Preparation of Platinum ethylenediamine-5'-acetamidofluorescein-nitrate.

5 [Pt(en)(5'-acetamidofluorescein)(NO₃)]NO₃.

All reactions are performed in the dark.

Dissolve Pt(en)(NO₃)₂ (0.047 mmol, 17.8 mg) in 5 ml methanol, heat until dissolving.

10 Dissolve 5' acetamidofluorescein (0.049 mmol, 20 mg, Molecular Probes Inc.) in 10 ml methanol.

Add the acetamidofluorescein solution to the platinum solution and react for at least 3 hours

Isolate end product by rotation evaporation.

15

Example 6

Preparation of Platinum ethylenediamine ε-(6-(fluorescein-5-carboxamido)hexanoyl)-L-lysine-nitrate.

20 [Pt(en)Fluoresceinlysine(NO₃)]NO₃.

All reactions are performed in the dark.

Dissolve Pt(en)(NO₃)₂ (0.077 mmol, 29.17 mg) in 10 ml methanol, heat upon dissolving.

25 Dissolve ε-(6-(fluorescein-5-carboxamido)hexanoyl)-L-lysine (0.081 mmol, 50 mg) in 10 ml methanol.

Add the fluorescein solution to the Platinum solution and react for at least 3 hours. Isolate the end product by rotation evaporation.

30

Example 7

Preparation of Platinum ethylenediamine-5-(and 6)-((N-(5-aminopentyl) amino) carbonyl) tetramethylrhodamine-nitrate.

35 [Pt(en)(tetramethylrhodamine cadaverine)(NO₃)]NO₃

All reactions are performed in the dark.

40 Dissolve Pt(en)(NO₃)₂ (0.074 mmol, 28 mg) in 10 ml methanol, heat upon dissolving.

Dissolve tetramethylrhodamine cadaverine (0.077 mmol, 40 mg, Molecular Probes Inc.) in 10 ml methanol.

Add the two solutions together and react for at least 3 hours.

5 Isolate the end product by rotation evaporation.

Example 8

10 Preparation of Platinum ethylenediamine-Cascade Blue^R cadaverine-nitrate.
[Pt(en)(Cascade Blue cadaverine)(NO₃)]NO₃.

Dissolve Pt(en)(NO₃)₂ (0.057 mmol, 21,6 mg) in 10 ml demineralised water and heat to 50°C until Pt(en)(NO₃)₂ is
15 completely dissolved.

Dissolve Cascade Blue cadaverine (0.06 mmol, 40 mg., Molecular Probes Inc.) in 10 ml demineralised water.

Add the two solutions together and react for at least 3 hours.

20 Isolate the end product by freeze-drying.

Cascade Blue^R is a registered trademark of Molecular Probes Inc.

25 Example 9

Preparation of
Platinum ethylenediamine-4,4-difluoro-5,7-dimethyl-4-boraO3a,4a-diaza-s-indacene-3-
30 propionylethylenediamine-nitrate.
[Pt(en)(BODIPY530/550 C3EDA)(NO₃)]NO₃.

Dissolve Pt(en)(NO₃)₂ (0.052 mmol, 19.66 mg) in 10 ml methanol and heat until dissolving .

35 Dissolve BODIPY 530/550 C3EDA (0.0545 mmol, 256 mg, Molecular Probes Inc.) in 10 ml methanol.

Add the two solutions together and react for at least 3 hours.

Isolate the end product by rotation evaporation.

40 BODIPY[®] is a registered trademark of Molecular Probes Inc.

Example 10

Preparation of Platinum ethylenediamine-N-ε-(5-
5 dimethylaminonaphthalene-1-sulfonyl)-L-lysine-nitrate
[Pt(en)(Dansyl lysine)(NO₃)]NO₃.

Dissolve Pt(en)(NO₃)₂ (0.125 mmol, 47.5 mg) in 20 ml
demineralised water and heat to 50°C until Pt(en)(NO₃)₂ is
10 completely dissolved.

Dissolve Dansyl lysine (0.13 mmol, 50 mg, Molecular Probes
Inc.) in 20 ml demineralised water, adjust the pH to 7 - 8
with 0.4 M NaOH.

Add the two solutions together and react for at least
15 3 hours. Isolate the end product by freeze-drying.

Example 11

Reaction for coupling Pt(en)-compounds to DNA
20

Typical reaction for labelling DNA molecules with a
Pt(en)-compound.

5 µg of double stranded DNA is sonicated or DNase treated
25 to yield fragments of 300-500 bp. 6 µg of Pt(en)-compound is
added and the volume is adjusted to 50 µl with demineralised
water. The reaction mixture is incubated at 65°C for 1 hour.
Non-bound Pt(en)-compound is blocked by adding 100 µl of a
NADDTTC solution. The Pt(en)-compound labelled DNA is purified
30 on a sephadex G-50 column. Readily labelled and purified DNA
is stored at -20°C or used directly in a DNA probe based
assay. Pt(en)-compound labelled DNA probes can be stored at
least 2 years at 20°C without loss of activity and/or
specificity.

35 All applications mentioned are carried out with probes
labelled according to this protocol.

Example 12

Preparation of Platinum ethylenediamine-rhodamine 123-nitrate.

5 [Pt(en)(rhodamine 123)(NO₃)]NO₃.

All reactions are performed in the dark.

Dissolve Pt(en)(NO₃)₂ (0.058 mmol, 21,9 mg) in 10 ml N,N-dimethylformamide, heat upon dissolving.

10 Dissolve rhodamine 123 (0.060 mmol, 22,8 mg) in 8 ml N,N-dimethylformamide. Add the rhodamine solution to the platinum solution and react for at least 3 hours. Isolate the product by rotation evaporation.

15 Example 13

Preparation of Platinum ethylenediamine-(7-amino-4-methylcoumarine)-nitrate.

[Pt(en)(7-amino-4-methylcoumarine)(NO₃)]NO₃.

20

All reactions are performed in the dark.

Dissolve Pt(en)(NO₃)₂ (0.098 mmol, 37,0 mg) in 25 ml methanol/2 ml demineralized water, heat upon dissolving.

25 Dissolve 7-amino-4-methylcoumarine (0.10 mmol, 18 mg) in 10 ml methanol.

Add the 7-amino-4-methylcoumarine solution to the platinum solution and react for at least 4 hours. Isolate the product by rotation evaporation, followed by freeze-drying.

30 Example 14

Preparation of Platinum ethylenediamine-5-aminoeosin-nitrate.

[Pt(en)(5-aminoeosin)(NO₃)]NO₃.

35

All reactions are performed in the dark.

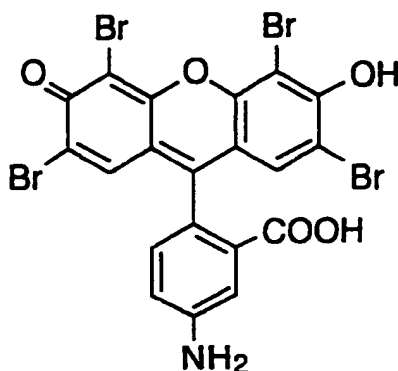
Dissolve Pt(en)(NO₃)₂ (0.068 mmol, 25.6 mg) in 15 ml N,N-dimethylformamide, heat upon dissolving.

Dissolve 5-aminoeosine (0.075 mmol, 50 mg) in 10 ml N,N-dimethylformamide.

Add the 5-aminoeosine solution to the platinum solution and react for at least 4 hours.

- 5 Isolate the product by rotation evaporation.

5-aminoeosin: commercially available by
Mol. Probes (A-117)



10

Example 15

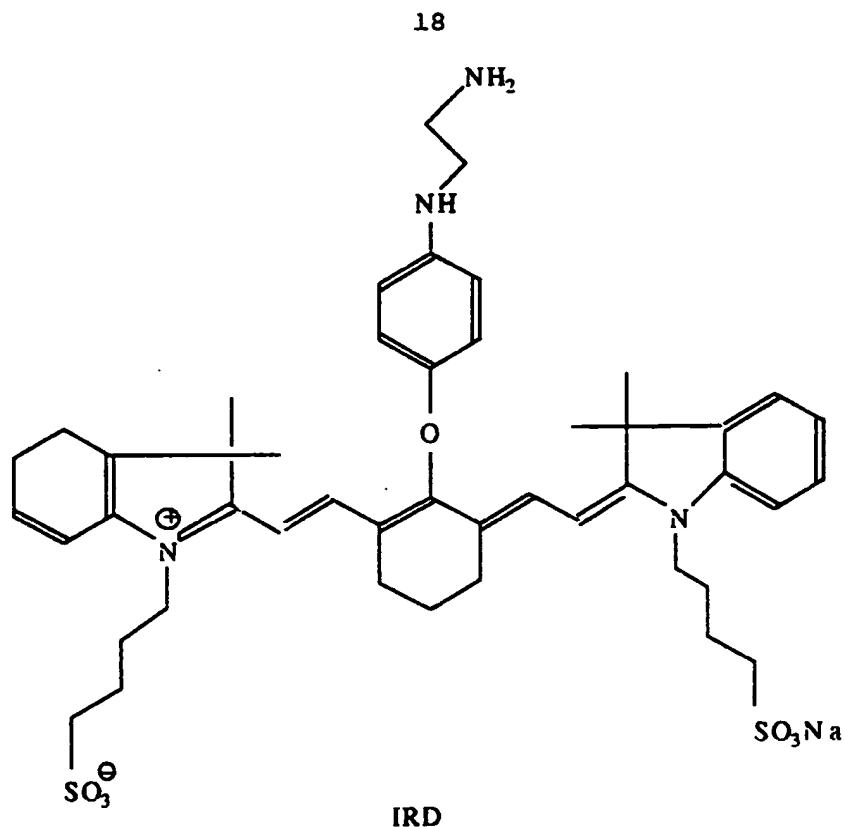
- 15 Preparation of Platinum ethylenediamine-IRD-nitrate.
[Pt(en)(IRD)(NO₃)]NO₃.

All reactions are performed in the dark.

Dissolve Pt(en)(NO₃)₂ (0.008 mmol, 3.2 mg) in 5 ml N,N-dimethylformamide, heat upon dissolving.

- 20 Dissolve IRD (presented below) (0.009 mmol, 7.5 mg) in 5 ml N,N-dimethylformamide.

Add the IRD solution to the platinum solution and react for at least 3 hours. Isolate the product by rotation evaporation.



Example 16

5 BIOTIN LABELLING OF DNA PROBES WITH THE UNIVERSAL LINKAGE SYSTEM

Introduction

10 The labelling method has been used to label DNA probes with biotin for In Situ Hybridization (ISH). In this example labelling procedures including the protocols and data for quality control procedures, are presented. For biotin labelling a plasmid cloned total DNA of Human Papilloma Virus type 6 (HPV-6, 40% GC basepairs) was used.

Experimental procedures

Plasmid DNA preparation

Total DNA of Human Papilloma Virus type 6 was cloned into
5 vector pSp-64. Plasmid DNA was transfected into E.coli (C-600)
and plated onto ampicillin containing LB plates. Single
colonies were grown overnight in large culture.
Plasmid DNA was isolated according to the method of Birnboim
and Doly¹, purified by Sepharose Cl-2B column chromatography
10 (Pharmacia) and checked for inserts by restriction-enzyme-
analyses. Plasmid DNA concentration was determined by A₂₆₀/280
absorption. After ethanol precipitation the DNA was
reconstituted in 10mM TRIS/HCl pH 7.2, 0.3mM EDTA to a final
concentration of 1 µg/µl (batch# 150894) Subsequently this DNA
15 was sonicated (Soniprep 150., MSE) for 3 times 10 minutes
(amplitude 5 microns) on ice.

The size of the resulting DNA fragments was determined
by 2% agarose gel electrophoresis and found to be in between
200-400 basepairs (batch# 051094).

20

Plasmid DNA labelling and purification

Plasmid HPV-6 DNA was labelled with Biotin-ULS by mixing
the following reagents:

plasmid HPV-6 DNA (batch# 051094)	5 µl (1 µg/µl)
25 Biotin-ULS labelling reagent (batch# BX940830)	8 µl (1 µg/µl)
Demineralised water (< 0.2/ µS/cm)	37 µl

The 50 µl reaction mixture was incubated for 15 minutes
at 85°C.

30 Excess of labelling reagent was captured by adding 50 µl
sodium diethyldithiocarbamate (2% solution in demineralised
water) and incubating for 30 minutes at room temperature.

Unbound Biotin-ULS was removed, using a S300 HR microspin
column (Pharmacia), by size exclusion chromatography.

35 Eluent volume was adapted to 500 µl giving a 10 ng/µl
biotin HPV-6 probe concentration (batch-06109-4.).

Quality Control for detection limits

The detection limit of the Biotin probe was determined by direct spot-blot and reversed filterhybridization according to the following protocols:

5

Direct spot blot

Biotin labeled HPV-6 probe (batch 061094) was 10 fold serially diluted into spot buffer comprising 900mM sodiumchloride, 90mM sodiumcitrate and 200 µg/ml single stranded salmon sperm DNA giving a dilution series varying from 1000 - 0.1 pg biotin probe per µl.

1 µl spots were applied onto Nitrocellulose membrane and incubated for 2 hours at 80°C to bind the DNA. The biotin probe was visualized using a streptavidin-alkaline phosphatase conjugate (Sigma) combined with an NBT/BCIP precipitating substrate solution (Sigma) according to the following protocol:

- Membranes were soaked in TBS solution containing 0.5% tween20 (TBST) for 5 minutes.
- 20 - Membranes were incubated with Strep-AP (3 DEA U/ml) in TBST for 15 minutes at 37°C.
- NC-membranes were washed 3 times 5 minutes in TBS solution followed by a 5 minutes wash step in demineralized water.
- Membranes were incubated with NBT/BCIP substrate solution
- 25 for 15 minutes at 37°C, subsequently washed in demineralized water and air dried.

Results

By using this method the detection limit of the biotin DNA probe was found to be less than 1 pg.

Reversed filterhybridization

HPV-6 DNA (batch 051094) was 1 in 10 diluted in 0.1N NaOH, incubated at 100°C for 5 minutes and directly placed on ice for 5 minutes to make the DNA single stranded.

A 10 fold serial dilution was made in cold 0.1N NaOH to give a serie varying from 10,000 - 1 pg DNA per µl. 1 µl spots were

applied onto Nylon membrane (Boehringer Mannheim) and air dried.

Biotin labelled HPV-6 DNA probe was diluted in 5xSSPE 0.5% SDS solution to yield a concentration of 200 ng/ml.

- 5 This solution was incubated for 5 minutes at 100°C and placed directly on ice for 5 minutes.

Nylon membranes containing target DNA were soaked in 2x SSC for 5 minutes and subsequently incubated with the single stranded probe solution for 2 hours at 37°C.

- 10 Excess of the biotin probe was removed by three changes in 2x SSPE 0.1% SDS for 10 minutes at 37°C followed by a 5 minutes TBST incubation.

The Biotin probe was visualized by performing the same protocol as described in the direct spotblot method.

15

Results

By using this procedure the detection limit of the biotin probe was found to be less than 10 pg.

20 Performance in In Situ Hybridisation

The test material consisted of 6 µm paraffin sections of a HPV-6 positive cervical condyloma mounted on organosilane coated glass slides.

- 25 The following protocol was applied (unless otherwise stated steps are at roomtemperature):

1 Paraffin sections were dewaxed in 3 changes of xylene and hydrated in graded ethanol.

2 Sections were rinsed in TBST for 5 minutes.

- 3 Sections were digested in 0.25% pepsin in 0.1N HCl for
30 30 minutes at 37°C, dehydrated in graded ethanol and air dried.

4 10 µl of probe solution was applied to a section and covered with a coverslip.

- 35 Probe solution consisted of biotin HPV-6 probe DNA (batch# 061094) in a concentration of 2 ng/µl dissolved in hybridization mixture comprising 0.6M NaCl, 0.06M Na-citrate, 35% formamid,

10% dextran sulphate, 2.5x Denhardt's and 10 µl/ml single stranded salmon sperm DNA.

- 5 Slides were placed on a hotplate set at 95°C for 5 minutes to denature probe and target DNA simultaneously.
- 5 6 Hybridisation was performed by placing the slides in a humidified chamber at 37°C for 2 hours.
- 7 Coverslips were removed and slides were washed in 3 changes of 15mM NaCl, 1.5mM Na-citrate and 5% formamide for 10 minutes at 37°C.
- 10 8 Slides were rinsed in TBST.
- 9 Sections were incubated with Streptavidin AP conjugate (3 DEA U/ml in TBST) for 15 minutes at 37°C.
- 10 Slides were washed in TBST (3x) and demineralised water (1x) for 5 minutes.
- 15 11 Sections were incubated with NBT/BCIP substrate solution for 15 minutes at 37°C.
- 12 Slides were washed in demineralised water (3x) for 1 minute and sections were mounted in glycerol/gelatin.

20 Results

By using the sections showed blue/purple precipitates at the sites of HPV-6 infected cells and minor background in the remaining tissue.

25 Conclusions

- The results demonstrate that DNA labelled with the ULS method have good label detection limits. The ULS method is very well suited for research, routine and for industrial production of labelled nucleic acids, as the method is fast and easy to perform, very sensitive, and does not include any enzymatic step, which makes it highly reproducible and fitted for an overall low cost production. The ULS method offer a useful alternative equaling conventional non-isotopic labelling methods.
- 30

General references

1. Maniatis T., Sambrook J., Fritsch E.F., Molecular Cloning, Second Edition, Cold Spring Harbor Laboratory Press, ISBN 0-87969-309-6.
- 5 2. Keller G.H., Manak M.M., DNA probes, Stockton Press, ISBN 0-333-47659-X.

Applications

- 10 1. The use of Pt-DNA linkers in the so called LIDIA technique: Linked DNA immuno Assay.

The LIDIA technique enables the quantitative analysis of small amounts of DNA (or RNA) e.g. after a PCR amplification
15 of the starting material. The technique is sensitive and specific, due to the use of specific DNA(RNA) probes and easy to perform, because of the quick DNA(RNA) Pt-labelling steps.

Description of the technique:

20

The technique uses fast Pt labelling compounds to label DNA(RNA) probes

This technique is possible with 3 different approaches.

- 25 1. Linking DNA probes molecules to a surface by using a Pt compound which cross-link DNA molecules irreversibly to plastic, nylon or nitrocellulose. Detection of DNA targets can then be accomplished by using classically labelled DNA/RNA probes. (nick translation or chemical modification,
30 random priming)
2. Linking a detectable group to the DNA, to render a DNA molecule into a so-called DNA probe. Binding of DNA compounds to a surface can then be accomplished by using classic techniques known to science (covalent linking to
35 specially treated microtiter plates, baking of DNA molecules onto nitrocellulose or binding of DNA molecules to nylon membranes.
3. A combination of techniques 1 and 2

Approach 1

An immobilized DNA probe can be used to catch specific target molecules in a sample by using a hybridization technique. Detection of formed hybrids can be done by using different techniques, e.g. a second labelled DNA probe can be used to hybridize with a different site on the target DNA molecule to form a sandwich hybrid. The label can then be detected by using state of the art immunological detection and colouring techniques.

Approach 2

A volume containing (amplified) detectable DNA(RNA) is directly labelled according to the protocol.

Excess label is quenched by adding NaDDTC or Thioureaum. This approach distinguishes itself from other techniques by the fact that the target molecule is labelled in contrast to other assay were labelled DNA(RNA) probes are used to detect the target. The quick binding capacity of the Pt-label compound(ULS) enables a DNA binding step as a routine step in a diagnostic test procedure (normal binding times are 60 minutes at 65°C).

A second step is performed in a microtiter plate precoated with a target specific probe. Incubation is allowed to the formation of stable "Labelled target" and probe hybrids. The direct labelling of target molecules enables the omission of laborious double hybridization techniques where one probe is used to catch the target and another labelled probe is used to detect the immobilized target.

In this method the probes are covalently linked to the microtiter plate to the surface of the wells. The second incubation step has the character of a liquid hybridisation and therefore can be performed very rapidly. This is one of the main innovative features of this approach to quantitative DNA hybridisation techniques.

Approach 3

Both for the immobilization of DNA probes or DNA targets and for the labelling of DNA probes and targets the newly developed Pt Universal Labelling System can be used. These two DNA linking techniques can be combined into one assay where both the "catcher" and the "detector" are linked to a second substance (either a detectable group like biotin, digoxigenin or a carrier surface like a plastic stick, microtiter plate or a membrane).

Examples of the technique: the detection of STD related microorganisms in human diagnostics (Chlamydia, Syphilis, AIDS, Herpes, Gonorrhoea, Hep. B,)

2. The use of Pt-DNA labels in combination with test strip procedures and formats.
The "DNA Dipstick".

The DNA dipstick technique enables the qualitative and semi-quantitative analysis of small amounts of DNA (or RNA) e.g. after a PCR amplification or freely present in samples of body fluids (blood, urine, sweat etc.)

The technique is sensitive and specific, due to the use of specific DNA (RNA) probes and easy to perform because of the quick DNA (RNA) Pt-labelling steps.

The universal labelling characteristics of the newly developed Pt label can be used in 3 ways to achieve a bound DNA (RNA) molecule.

1. It can be used to attach a detectable marker group to a polynucleotide sequence.
2. It can be used to attach polynucleotide sequences irreversibly to a solid phase (plastic, membranes, latex beads, hydrosols or microtiter plate wells).

3. A combination of 1 and 2

ad 1:

In this example there is a twofold approach to the detection of biological analytes in test samples.

5 Firstly a DNA probe can be labelled with the newly developed Pt labelling compound. This labelled probe can then be used to detect preformed hybrids on a membrane formed between the target DNA sequence and a primary probe. It is essential in this method that the primary probe recognizes a different
10 sequence on the target than the secondary Pt labelled probe. In practice, this can be achieved for instance with RNA hybridization where a POLY A probe is used as a primary probe to immobilize all RNA (recognizable by its polyT tails) to a membrane.

15 The second approach differs slightly in that in this case the target can be labelled in the test sample fluid, because of the fast and very specific Pt labelling characteristics. A procedure like this would comprise a catch of the labelled target with an immobilized specific unlabelled DNA probe on a
20 suitable membrane. Hence a dipstick version for DNA/RNA applications.

ad 2:

To immobilize DNA probes or target DNA, a non-labelled Pt
25 compound (that is a Pt compound with 2 free binding sites) can be used to act as a bridge between DNA and the surface of carriers (plastic, membrane, microtiter plates etc.)

It greatly enhances the usability of DNA sequences as catcher molecules in diagnostic assays, since there are little
30 substances known to science that bind readily DNA in a spontaneous way. Introducing this Pt bridge molecule a wide field of new applications for the DNA technology has come within reach.

35 ad 3: a combination of example 1 and 2

General: especially the use of the Pt linker molecule in latex or hydrosol assays is particularly interesting. The

linker enables the coupling of DNA molecules to small particles. The DNA molecules can be hybridized to target material. A positive reaction is visualized by an agglutination of the particles, due to crosslinking of the DNA hybrid particle compounds.

A test like this can be made quantative, the rate of agglutination can be tuned and measured at a specific wavelength. Especially gold particles have the intrinsic characteristic that a shift in optimal wavelength occurs after agglutination.

3. Protein labelling using Pt-marker compounds in diagnostic test procedures.

The ability of Pt-marker compounds to interact with protein structures under specified conditions that differ from the DNA binding conditions, can be used to label e.g.:

- a. mono or polyclonal antibodies to detect primary targets in test samples
- b. protein derivatives used as antigens in test procedures based on the so called inhibition principle
- c. specific protein structures that are known to interact specifically with other compounds, but generate a non immunogenic interaction with each other. e.g. streptavidin and biotin, and protein A and G with Immunoglobulines.

Labelled tracer molecules can be used to determine either qualitatively or quantitatively an amount of a certain biolyte in a suitable test format (dipsticks, elisa or others)

4. Long term acting proteins:

Of special interest is the potential use of the Pt linker to stabilise protein (biomolecules in general) in vivo. A therapeutical use of the Pt compound is that bio-substances used as therapeutics (insulin, factor VIII and the like) show a short term action once inside the body. The coupling of these substances to Pt compound may enhance the halflife of

these molecules thus enabling longer acting substances to be produced and used in vivo. This implies less trouble for patients and more effective use of expensive drugs and medicines.

5

5. Detection of Platinated DNA probes with the the silver-enhancement technique.

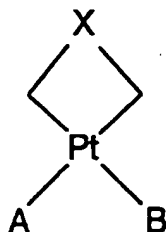
Platinated DNA/RNA probe can be employed in hybridisation methods to detect DNA/RNA sequences in sample material. The introduction of a platinum compound at the site of the target enables the deposition of Ag molecules in a chemical reaction especially designed to reduce ionic Silver to metallic silver. At the site of a Pt nucleus a decomposition of metallic Silver(black) occurs due to the catalytic effect of the Pt nucleus.

Ionic silver is reduced by a reducing agent (e.g. Na-borohydrid, Hydrochinon) in solution. In a constant ratio the amount of silver deposited on the Pt is proportional to the length of the enhancement incubation. Visualisation of a non-visible Pt nucleus can be accomplished by the empirical observation of the appearace of a black spot in the test sample.

The black spots indicate the site of specific probes binding and thus the site of specific target location. The technique enables a quick and easy diagnostic procedure for the detection of various microorganisms and gene translocations/abnormalities.

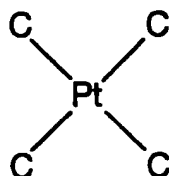
CLAIMS

1. Method for the preparation of linkers for labelled substances, whereby the linker comprises a platinum compound of the formula:



wherein X represents any stabilizing bridge and wherein A and B represent the same or different reactive moieties,

10 whereby the starting material comprises a compound of the structure:



15 wherein C represents an electronegative reactive moiety.

2. A method according to claim 1 wherein A and B are the same.

20 3. A method according to claim 1-2, wherein X represents an alifatic diamine.

4. A method according to claim 3 wherein X represents a diamine having 2-6 carbon atoms.

25

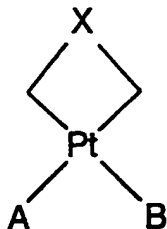
5. A method according to claim 2 wherein X is an ethylene diamine group.

6. A method according to claim 5 wherein A and B are selected from the group consisting of NO_3^- , SO_3^- , Cl^- , I^- , or other halogens.

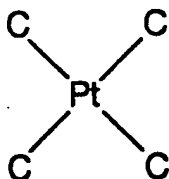
5

7. A method according to anyone of the afore going claims wherein C is a halogen, NO_3^- or SO_3^- .

8. A method for the preparation of linkers for labelled
10 substances, whereby the linker comprises a platinum compound of the formula:



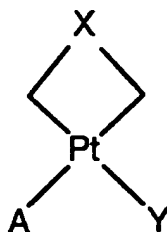
15 wherein X represents an ethylene diamine group and wherein A and B represent NO_3^- , whereby the starting material comprises a compound of the structure:



20

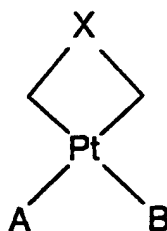
wherein C represents a halogenic group, wherein the starting compound is reacted with ethylene diamine and whereby the resulting compound is reacted with AgNO_3 .

9. A method for preparing a labelling substance of the formula:



5

wherein X and A are as defined in the afore going claims and wherein Y is a label, whereby a compound of formula:



10

wherein A,B and X are defined as in the foregoing claims is reacted with a labelling group whereby Y is substituted for B.

10. A method for labelling a bio-organic molecule whereby a labelling compound obtainable by the method of claim 9 is reacted with the bio-organic molecule, whereby the group A is replaced by the bio-organic molecule.

11. Method according to claim 10 whereby the bio-organic molecule is a protein, a peptide, a DNA molecule or an RNA molecule, or an (oligo-)nucleotide.

12. A diagnostic test kit for detecting, determining and or localizing biological substances of interest, comprising a linker obtainable according to the methods of any one of claims 1-8 together with at least one suitable label and optionally other suitable means for detection.

13. A diagnostic test kit for detecting, determining and or
localizing biological substances of interest, comprising a
labelling substance obtainable according to the method of
claim 9, optionally together with other suitable means for
5 detection.

14. diagnostic test kit for detecting, determining and or
localizing biological substances of interest, comprising a
labelled bio-organic molecule obtainable according to the
10 methods of claim 10 or 11, optionally together with other
suitable means for detection.

INTERNATIONAL SEARCH REPORT

International Application No

PCT/NL 96/00198

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C07F15/00 G01N33/58

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C07F G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	GB,A,2 148 891 (NIPPON KAYAKU KABUSHIKI KAISHA) 5 June 1985 see claim 16 ---	8
X	PATENT ABSTRACTS OF JAPAN vol. 5, no. 180 (C-79) [852] , 19 November 1981 & JP,A,56 103192 (YOSHINORI KITANI), 18 August 1981, see abstract ---	8
Y	EP,A,0 386 243 (SAGAMI CHEMICAL RESEARCH CENTER) 12 September 1990 see example 5 ---	8
Y	EP,A,0 282 672 (NIPPON KAYAKU KABUSHIKI KAISHA) 21 September 1988 see claim 7 ---	8
-/--		



Further documents are listed in the continuation of box C.



Patent family members are listed in annex.

* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- "&" document member of the same patent family

Date of the actual completion of the international search

5 August 1996

Date of mailing of the international search report

20.08.96

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Authorized officer

Kapteyn, H

INTERNATIONAL SEARCH REPORT

International Application No

PCT/NL 96/00198

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	US,A,4 207 416 (J.D. HOESCHELE) 10 June 1980 see claim 1	9,10
A	--- WO,A,92 01699 (STICHTING KLINISCHE RESEARCH ACADEMISCH ET AL) 6 February 1992 cited in the application * complete document * -----	9,10

INTERNATIONAL SEARCH REPORT

International application No.

PCT/NL96/00198

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☒ Claims Nos.: 1-7
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
The preparation method of claims 1 to 7 are incomplete. It is not clear under which conditions the starting materials are converted to the claimed linkers.

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.

2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

☐ The additional search fees were accompanied by the applicant's protest.

☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/NL 96/00198

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
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		US-A- 4861905	29-08-89
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